

# Laminin-5 $\gamma$ 2 Chain and Matrix Metalloproteinase-2 Expression in the Neoplastic Changes of Uterine Cervical Squamous Epithelium

ROBIN MALINA, SATORU MOTOYAMA, SHINYA HAMANA  
and TAKESHI MARUO

*Department of Obstetrics and Gynecology,  
Kobe University Graduate School of Medicine, Kobe, Japan*

Received 15 January 2005 /Accepted 15 April 2005

**Key words:** laminin-5  $\gamma$ 2 chain, matrix metalloproteinase-2, uterine cervical neoplasia, immunohistochemistry

Laminin-5 is an essential component of epithelial basal membranes. Matrix metalloproteinase-2 (MMP-2) changes laminin-5 from static to migratory substrate by cleaving the short arm of the  $\gamma$ 2 subunit of the laminin-5. Recently, the expression of laminin-5  $\gamma$ 2 chain has been reported to be a marker of invasiveness of epithelial malignant tumors. We investigated the expression of laminin-5  $\gamma$ 2 chain and MMP-2 in the neoplastic changes of uterine cervical squamous epithelium. Tissue samples of normal uterine cervix, cervical intraepithelial neoplasia-1, -3 (CIN-1, -3) and squamous cell carcinoma keratinizing (SCC-K) and non-keratinizing (SCC-NK) were analyzed for expression of the laminin-5  $\gamma$ 2 chain and MMP-2 by immunohistochemistry. In normal uterine cervix and CIN-1, laminin-5  $\gamma$ 2 chain was immunolocalized only in the basal membrane. In CIN-3, expression of laminin-5  $\gamma$ 2 chain and MMP-2 became apparent at the epithelial-stromal interface of epithelial cells. In SCC-K and SCC-NK, laminin-5  $\gamma$ 2 chain and MMP-2 expression were abundant at the epithelial-stromal interface of tumor clusters. These results suggest that the expression of laminin-5  $\gamma$ 2 chain and MMP-2 may be involved in the process of neoplastic changes of uterine cervical squamous epithelium.

## INTRODUCTION

The group of laminins is composed of extracellular matrix adhesion molecules localized to the basement membrane in most tissues (6,16). Laminin-5 consists of three subunits; one heavy  $\alpha$ 3 and two light,  $\beta$ 3 and  $\gamma$ 2 chains (5,27,36). The laminin-5 isoform, which is an essential component of epithelial basement membrane, is involved in cell anchorage, tissue remodeling, wound healing and cancer invasion (7,20,28,30,32). Like other laminins, laminin-5 interacts with cell surface receptors. Laminin-5 is a recognized ligand for integrins  $\alpha$ 6 $\beta$ 4,  $\alpha$ 3 $\beta$ 1 and  $\alpha$ 6 $\beta$ 1 (5,21). Several studies have shown that laminin-5 is frequently expressed at the invasive margin (tumor budding) of colorectal (25,26,32), gastric (14), pancreatic (31), breast adenocarcinomas (25), uterine cervical (25,26,30) and oral (9,15,23) squamous cell carcinomas. Laminin-5 promotes the adhesion, migration, and scattering of various types of cultured cells more strongly than other known extracellular matrix proteins (10,11,19). These activities are mediated mainly by integrin  $\alpha$ 3 $\beta$ 1 (5,11,38). However, the regulatory mechanism for the overexpression of laminin-5 in cancer is currently unclear.

Matrix metalloproteinase-2 (MMP-2) which cleaves a number of substrates including native types I, IV, V, VII, and XI collagen, denatured collagens, elastin, proteoglycans, laminin and fibronectin, is a member of the MMP family of zinc-dependent endopeptidases (3,4,18,34,37). MMP-2 activation is associated with the expression of tissue-invasive phenotypes (1,4,22,35). MMP-2 changes laminin-5 from static to migratory substrate by cleaving the short arm of the  $\gamma 2$  subunit of the laminin (7). MMP-2 is thought to play an important role in the degradation of extracellular matrix, which is an essential step in the cascade of metastasis and angiogenesis.

In the present study, we immunohistochemically examined the expression of laminin-5  $\gamma 2$  chain and MMP-2 in the neoplastic changes of uterine cervical squamous epithelium.

## MATERIAL AND METHODS

### Tissue samples

After informed consent was obtained from each patient, stored tissues from the Department of Pathology, Kobe University Hospital, Kobe, Japan were used in this study. The specimens of patients diagnosed between 1999 and 2002 were formaldehyde fixed, paraffin embedded and diagnosed on hematoxylin-eosin (H-E)-stained tissue sections. Tumors were classified according to the WHO classification.

The tissue samples used consisted of 5 normal uterine cervical tissues, 5 CIN-1, 5 CIN-3, 5 SCC-K and 5 SCC-NK. Five-micron-thick paraffin sections were used for immunohistochemical staining. Invasiveness was graded according to the International Federation of Gynecology and Obstetrics (FIGO) classification system as follows: carcinoma in situ (FIGO 0), microinvasive cancer (FIGO IA), and frankly invasive cancer (FIGO IB). The carcinoma samples included FIGO stages in a range IB1 to IIB SCC-K and age at diagnosis ranged from 27 to 68. The samples of SCC-NK included FIGO stages IA2 to IIB and age at diagnosis ranged from 45 to 75. All specimens were obtained before anti-tumor therapy was started.

### Immunohistochemistry

Immunohistochemical staining was carried out using the avidin-biotin immunoperoxidase technique (24) with the polyvalent immunoperoxidase kit (Omnitag, Lipshaw, MI, USA with DAB Chromogen). In order to improve the immunostaining efficacy, the antigen retrieval method was used (24). The sections were digested with Trypsin Sigma 0.2% for 30 minutes at 37°C (for laminin-5). Before deparaffinization, sections were heated for 1 hour at 37°C (for MMP-2). The sections were incubated with each antibody at 37°C for 90 minutes. Mouse anti-laminin 5 (laminin  $\gamma 2$  chain) monoclonal antibody (Chemicon Int.) at 5  $\mu$ l/ml dilution and anti-hMMP-2 purified mouse IgG monoclonal antibody F-68 (Daiichi Fine Chemical Co. L.T.D. against human) at 4  $\mu$ l/ml dilution were used as the primary antibody, respectively. The sections were counterstained with Mayer's Hematoxylin. The slides were dehydrated and mounted with a xylene-soluble mounting medium and examined microscopically. Control sections were subjected to the same technique except that the primary antibody was replaced by non-immune murine IgG at the same dilution as the specific antibody. No significant staining was observed in the negative controls.

Immunohistochemistry was analyzed as follows: negative (-) for no detectable immunostaining, ( $\pm$ ) for weak but definitely detectable immunostaining, (+) for moderate immunostaining, (++) abundant immunostaining. The immunostaining intensity was evaluated by two independent observers (R.M., S.H.).

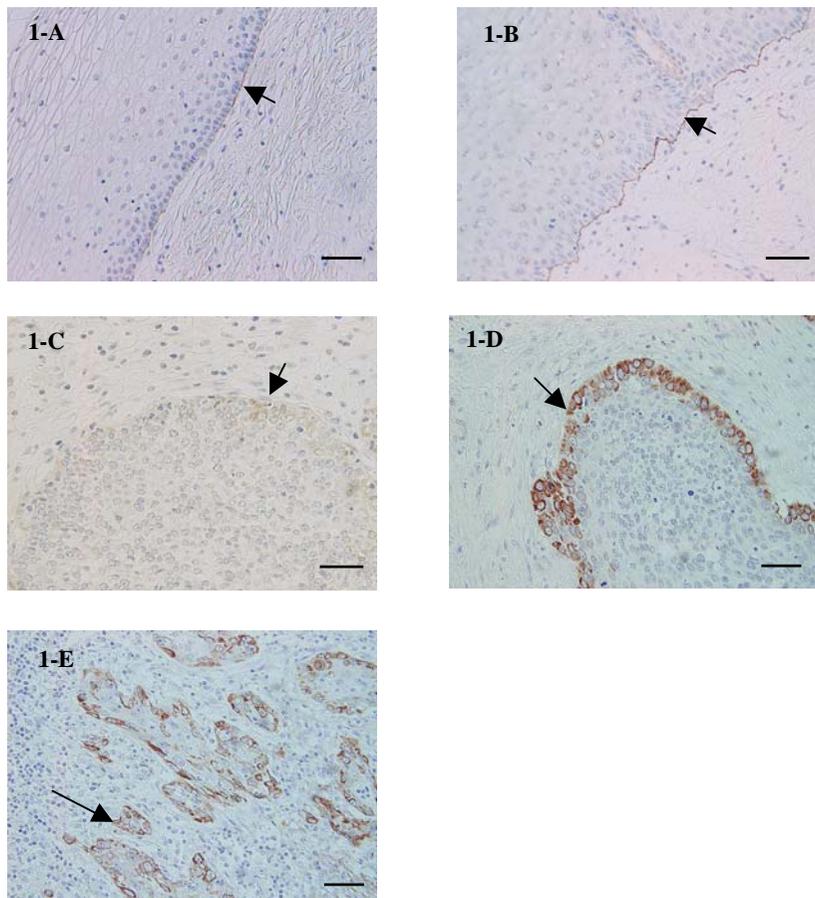
## LAMININ-5 $\gamma$ 2 CHAIN AND MMP-2 IN UTERINE CERVIX

### RESULTS

Immunohistochemical expression of laminin-5  $\gamma$ 2 chain in normal uterine cervical tissue and CIN-1 was noted only in the basement membrane. Basement membrane showed intense staining for the laminin-5  $\gamma$ 2 chain in a linear pattern. Peribasal and basal cells showed no positive immunostaining for laminin-5  $\gamma$ 2 chain (Fig.1-A, 1-B).

In CIN-3, weak to moderate immunostaining for laminin-5  $\gamma$ 2 chain was preferentially apparent in the cytoplasm of epithelial cells at the epithelial-stromal interface, whereas the basal membrane was negative for laminin-5  $\gamma$ 2 chain immunostaining (Fig.1-C).

SCC-K (Fig.1-D) showed a comparable expression for laminin-5  $\gamma$ 2 chain with SCC-NK (Fig.1-E). Immunostaining for laminin-5  $\gamma$ 2 chain was abundant in the cytoplasm of tumor cells localized at the cancer-stromal interface and in the tumor cells invading into the stroma, but negative in basal membrane in SCC-K and SCC-NK. The characteristics of neoplastic changes and results of the immunohistochemical analysis were summarized in Table 1.



**Figure 1.** Immunostaining for laminin-5  $\gamma$ 2 subunit in the uterine cervix. The basal membrane was positive for laminin-5  $\gamma$ 2 immunostaining in normal uterine cervix (Fig.1-A) and CIN-1 (Fig.1-B). In CIN-3, immunostaining for laminin-5  $\gamma$ 2 chain was weak to moderate in the epithelial cells at the epithelial-stromal interface, but negative in the basal membrane (Fig.1-C). In SCC-K (Fig.1-D) and SCC-NK (Fig.1-E), abundant cytoplasmic staining for laminin-5  $\gamma$ 2 chain was apparent at the epithelial-stromal interface and in invading tumor cells (tumor budding cancer cells), whereas the basal membrane was negative for the immunostaining. Arrows indicate the areas of positive immunostaining. (Bars represent 100  $\mu$ m. Original magnification, x200)

**Table 1.** Immunohistochemical staining for laminin-5  $\gamma$ 2 chain in the uterine cervix

|        | Basal Membrane | Epithelial-stromal Interface | Neoplastic Lesion   |
|--------|----------------|------------------------------|---|
| Normal | +              | -                            | No neoplastic cells   |
| CIN 1  | +              | -                            | Neoplastic cells involve less than 1/3 of the thickness of epithelium, koilocytotic changes |
| CIN 3  | -              | ±                            | 2/3 to full thickness epithelium replaced by neoplastic cells                               |
| SCC-K  | -              | ++                           | Carcinoma with keratinization   |
| SCC-NK | -              | ++                           | Carcinoma with no keratinization  |

Immunohistochemistry was analysed as follows: negative(-); weak(±); moderate(+); strong(++). Neoplastic cells show increased nuclear-cytoplasmic ratio, irregularity of nuclear size and nuclei

The same specimens were subjected to immunohistochemical analysis with anti-MMP-2 monoclonal antibody. In normal cervical tissue (Fig. 2-A) and CIN-1 (Fig.2-B) immunostaining for MMP-2 was negative at the basal membrane and basal cells, but positive in the suprabasal cells of the squamous epithelium. In CIN-3, moderate immunostaining for MMP-2 became apparent in the neoplastic cells (Fig. 2-C).

In invasive carcinomas, each subgroup of SCC-K and SCC-NK showed a distinct expression for MMP-2. SCC-K (Fig.2-D) showed diffuse staining pattern of MMP-2 in cytoplasm of cancer cells. In SCC-NK (Fig.2-E), the immunostaining for MMP-2 was abundant in the invasive front at the epithelial-stromal interface. The characteristics of neoplastic changes and results of the immunohistochemical analysis were summarized in Table 2.

**Table 2.** Immunohistochemical staining for MMP-2 in the uterine cervix

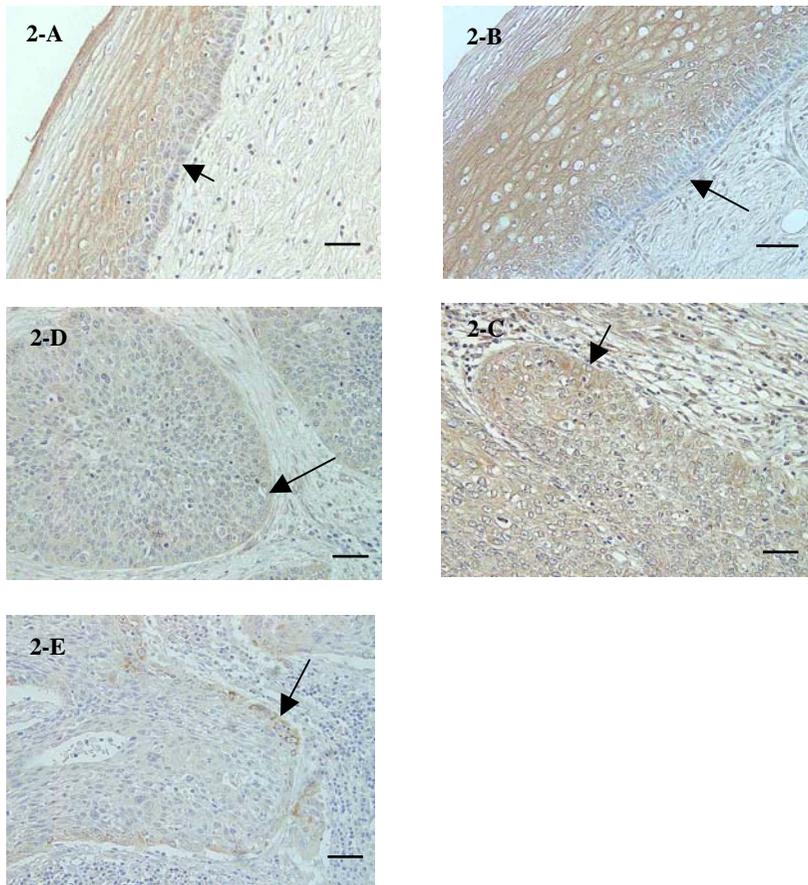
|        | Basal Membrane | Epithelial-stromal Interface | Neoplastic Lesion   |
|--------|----------------|------------------------------|---|
| Normal | -              | -                            | No neoplastic cells   |
| CIN 1  | -              | -                            | Neoplastic cells involve less than 1/3 of the thickness of epithelium, koilocytotic changes |
| CIN 3  | -              | +                            | 2/3 to full thickness epithelium replaced by neoplastic cells                               |
| SCC-K  | -              | +                            | Carcinoma with keratinization   |
| SCC-NK | -              | ++                           | Carcinoma with no keratinization  |

Immunohistochemistry was analysed as follows: negative(-); weak(±); moderate(+); strong(++). Neoplastic cells show increased nuclear-cytoplasmic ratio, irregularity of nuclear size and nuclear density.

## DISCUSSION

In the present study, immunohistochemical expression of laminin-5  $\gamma$ 2 chain and MMP-2 was found to increase according to the degree of neoplastic changes of uterine cervical epithelium. In normal uterine cervical tissues and CIN-1, the laminin-5  $\gamma$ 2 chain was

## LAMININ-5 $\gamma$ 2 CHAIN AND MMP-2 IN UTERINE CERVIX



**Figure 2.** Immunostaining for MMP-2 in the uterine cervix. The basal membrane in normal uterine cervix (Fig.2-A) and CIN-1 (Fig.2-B) was negative for immunostaining for MMP-2, whereas the suprabasal cells were positive for immunostaining for MMP-2. In CIN-3 (Fig.2-C), moderate immunostaining for MMP-2 was apparent in the neoplastic cells at the epithelial-stromal interface, but the basal membrane was negative for signal for MMP-2. In SCC-K (Fig.2-D), diffuse immunostaining for MMP-2 was noted in the tumor cells. In SCC-NK (Fig.2-E), cytoplasmic staining for MMP-2 was positive in the tumor cells at the epithelial-stromal interface and invading tumor cells (tumor budding cancer cells), whereas the basal membrane was negative for the immunostaining. Arrows indicate areas of positive immunostaining. (Bars represent 100  $\mu$ m. Original magnification, x200)

expressed along the basal membranes in a linear pattern. By contrast, the cytoplasmic immunoreactivity for laminin-5  $\gamma$ 2 chain was observed to be related with the degree of cervical intraepithelial neoplasia. These findings were partially different from previous study by Kohlbreger et al. (12) who showed no basal membrane staining in normal uterine cervix. A possible explanation could be the use of different immunohistochemical procedure with different sensitivity to laminin-5  $\gamma$ 2 chain. Skyldberg et al. (30) demonstrated no significant difference among expression of laminin-5  $\gamma$ 2 chain in CIN-1 and CIN-2 and consistent positive cytoplasmic expression of laminin-5  $\gamma$ 2 chain in invasive carcinomas. MMP-2 expression in normal uterine cervix and CIN-1 was positive in

suprabasal cells. This pattern of expression was similar to that in dysplastic epithelium of esophagus (29). Several studies reported that MMP-2 and laminin-5  $\gamma 2$  chain were involved in adhesion, migration and scattering of tumor cells and were associated with tumor budding (17). In invading human cancer cells, cytoplasmic staining for laminin-5 was apparent. The prognostic significance of this phenomenon was reported in colon adenocarcinoma. Hase *et al.* (8) found a positive correlation between the degree of tumor budding and recurrence of tumors following curative surgery. We have shown that laminin-5  $\gamma 2$  chain and MMP-2 were expressed at the cancer-stromal interface and at the invasive front of uterine cervical tumors.

The mechanism of the preferential expression of laminin-5  $\gamma 2$  chains at the invasive margin in cancer is not known yet. Koshikawa *et al.* found that  $\gamma 2$  chain of laminin 5 without a signal for  $\alpha 3$  and  $\beta 3$  was strongly expressed at the invasive margin of gastric carcinoma cells (14). These results suggest that laminin-5  $\gamma 2$  chain monomer may have a function different from the laminin-5 trimmer. On the other hand, Sordat *et al.* reported co-expression of the laminin  $\beta 3$  chain and  $\gamma 2$  chain in colorectal cancers (33). Predominant expression of the laminin  $\gamma 2$  chain at the invasive front has been reported in carcinomas of the colon, pancreas and stomach. Laminin-5 may act as a ligand for invading carcinoma cells. Gianneli *et al.* (7) have shown that MMP-2 changes laminin-5 from static to migratory substrate by cleaving the short arm of the  $\gamma 2$  subunit of the laminin-5. In the present study, increased expression of laminin-5  $\gamma 2$  chain and MMP-2 were noted at the cancer- stromal interface in SCC-NK, whereas in SCC-K, diffuse expression of MMP-2 was noted in the tumor cells.

The membrane-type 1 matrix metalloproteinase also plays an important role in cell migration on laminin-5 and cleaves laminin-5  $\gamma 2$  chain more efficiently than MMP-2 (13). It may be a reason why we found a different pattern of expression of MMP-2 between SCC-K and SCC-NK. Previous studies showed that MMP-2 was concentrated along the basement membrane at the sites of tissue remodeling (2) and at the leading edge of invading tumors (7). Masaki *et al.* (17) showed that although tumor cells might produce MMP-2, tumor cell budding was not associated with MMP-2 expression in the tumor cells, but it was associated with MMP-2 expression in stromal cells in colorectal carcinoma. Interactions of carcinoma cells with stromal cells or with the extracellular matrix surrounding cells at the invasive front may result in an accumulation of laminin-5  $\gamma 2$  chains and MMP-2 at the invasive front, where they may play a direct role in tumor invasion processes. Taking these findings into account, it is likely that laminin-5 and MMP-2 may be involved in the initial stage of tumorigenesis of uterine cervical squamous epithelium and thus the detection of laminin-5 and MMP-2 expression may be useful as a marker of invasiveness in uterine cervical lesions.

## REFERENCES

1. **Aimes, R.T., Quigley, J.P.** 1995. Matrix metalloproteinase-2 is an interstitial collagenase. Inhibitor-free enzyme catalyzes the cleavage of collagen fibrils and soluble native type I collagen generating the specific 3/4- and 1/4-length fragments. *J Biol Chem.* **270**:5872-6.
2. **Birkedal-Hansen, H.** 1995. Proteolytic remodeling of extracellular matrix. Review. *Curr Opin Cell Biol.* **7**:728-35.
3. **Brown, P.D., Bloxidge, R.E., Anderson, E., Howell, A.** 1993. Expression of activated gelatinase in human invasive breast carcinoma. *Clin Exp Metastasis.* **11**:183-9.
4. **Brown, P.D., Bloxidge, R.E., Stuart, N.S., Gatter, K.C. and Carmichael, J.** 1993.

## LAMININ-5 $\gamma$ 2 CHAIN AND MMP-2 IN UTERINE CERVIX

- Association between expression of activated 72-kilodalton gelatinase and tumor spread in non-small-cell lung carcinoma. *J Natl Cancer Inst.* **85**:574-8.
5. **Carter, W.G., Ryan, M.C. and Gahr, P. J.** 1991. Epiligrin, a new cell adhesion ligand for integrin alpha 3 beta 1 in epithelial basement membranes. *Cell.* **65**:599-610.
  6. **Engel, J.** 1992. Laminins and other strange proteins. Review. *Biochemistry.* **31**:10643-51.
  7. **Giannelli, G., Falk-Marzillier, J., Schiraldi, O., Stetler-Stevenson, W.G. and Quaranta, V.** 1997. Induction of cell migration by matrix metalloprotease-2 cleavage of laminin-5. *Science.* **277**:225-8.
  8. **Hase, K., Shatney, C., Johnson, D., Trollope, M., Vierra, M.** 1993. Prognostic value of tumor "budding" in patients with colorectal cancer. *Dis Colon Rectum.* **36**:627-35.
  9. **Kainulainen, T., Autio-Harmainen, H., Oikarinen, A., Salo, S., Tryggvason, K., Salo, T.** 1997. Altered distribution and synthesis of laminin-5 (kalinin) in oral lichen planus, epithelial dysplasias and squamous cell carcinomas. *Br J Dermatol.* **136**:331-6.
  10. **Kikkawa, Y., Akaogi, K., Mizushima, H., Yamanaka, N., Umeda, M., and Miyazaki, K.** 1996. Stimulation of endothelial cell migration in culture by ladsin, a laminin-5-like cell adhesion protein. *In Vitro Cell Dev Biol Anim.* **32**:46-52.
  11. **Kikkawa, Y., Umeda, M., and Miyazaki, K.** 1994. Marked stimulation of cell adhesion and motility by ladsin, a laminin-like scatter factor. *J Biochem.* **116**:862-9.
  12. **Kohlberger, P., Beneder, Ch., Horvat, R., Leodolter, S., Breitenacker, G.** 2003. Immunohistochemical expression of laminin-5 in cervical intraepithelial neoplasia. *Gynecol Oncol.* **89**:391-4.
  13. **Koshikawa, N., Giannelli, G., Cirulli, V., Miyazaki, K., and Quaranta, V.** 2000. Role of cell surface metalloprotease MT1-MMP in epithelial cell migration over laminin-5. *J Cell Biol.* 2000. **148**:615-24.
  14. **Koshikawa, N., Moriyama, K., Takamura, H., Mizushima, H., Nagashima, Y., Yanoma, S., Miyazaki, K.** 1999. Overexpression of laminin gamma 2 chain monomer in invading gastric carcinoma cells. *Cancer Res.* **59**:5596-601.
  15. **Kosmehl, H., Berndt, A., Strassburger, S., Borsi, L., Rousselle, P., Mandel, U., Hyckel, P., Zardi, L., Katenkamp, D.** 1999. Distribution of laminin and fibronectin isoforms in oral mucosa and oral squamous cell carcinoma. *Br J Cancer.* **81**:1071-9.
  16. **Malinda, K. M., and Kleinman, H. K.** 1996. The laminins. Review. *Int J Biochem Cell Biol.* **28**:957-9.
  17. **Masaki, T., Matsuoka, H., Sugiyama, M., Abe, N., Izumisato, Y., Goto, A., Sakamoto, A., Atomi, Y.** 2003. Laminin-5 gamma 2 chain and matrix metalloproteinase-2 may trigger colorectal carcinoma invasiveness through formation of budding tumor cells. *Anticancer Res.* **23**:4113-9.
  18. **Matrisian, L.M.** 1992. The matrix-degrading metalloproteinases. Review. *Bioessays.* **14**:455-63.
  19. **Miyazaki, K., Kikkawa, Y., Nakamura, A., Yasumitsu, H., and Umeda, M.** 1993. A large cell-adhesive scatter factor secreted by human gastric carcinoma cells. *Proc Natl Acad Sci U S A.* **90**:11767-71.
  20. **Nguyen, B.P., Ryan, M.C., Gil, S.G., and Carter, W.G.** 2000. Deposition of laminin 5 in epidermal wounds regulates integrin signaling and adhesion. Review. *Curr Opin Cell Biol.* **12**:554-62.
  21. **Niessen, C.M., Hogervorst, F., Jaspars, L.H., de Melker, A.A., Delwel, G.O., Hulsman, E.H., Kuikman, I., and Sonnenberg, A.** 1994. The alpha 6 beta 4 integrin is a receptor for both laminin and kalinin. *Exp Cell Res.* **211**:360-7.
  22. **Okada, A., Bellocq, J., Rouyer, N., Chenard, M., Rio, M., Chambon, P., and Basset, P.** 1995. Membrane-type matrix metalloproteinase (MT-MMP) gene is expressed in stromal cells of human colon, breast, and head and neck carcinomas. *Proc Natl Acad*

- Sci U S A. **92**:2730-4.
23. **Ono, Y., Nakanishi, Y., Ino, Y., Niki, T., Yamada, T., Yoshimura, K., Saikawa, M., Nakajima, T., Hirohashi, S.** 1999. Clinocopathologic significance of laminin-5 gamma 2 chain expression in squamous cell carcinoma of the tongue: immunohistochemical analysis of 67 lesions. *Cancer*. **85**:2315-21.
  24. **Paananen, R., Glumoff, V., Sormunen, R., Voorhout, W., and Hallman, M.** 2001. Expression and localization of lung surfactant protein B in Eustachian tube epithelium. *Am J Physiol Lung Cell Mol Physiol*. **280**:L214-20.
  25. **Pyke, C., Romer, J., Kallunki, P., Lund, L.R., Ralfkiaer, E., Dano, K., Tryggvason, K.** 1994. The gamma 2 chain of kalinin/laminin 5 is preferentially expressed in invading malignant cells in human cancers. *Am J Pathol*. **145**:782-91.
  26. **Pyke, C., Salo, S., Ralfkiaer, E., Romer, J., Dano, K., Tryggvason, K.** 1995. Laminin-5 is a marker of invading cancer cells in some human carcinomas and is coexpressed with the receptor for urokinase plasminogen activator in budding cancer cells in colon adenocarcinomas. *Cancer Res*. **55**:4132-9.
  27. **Rousselle, P., Lunstrum, G.P., Keene, D.R., and Burgeson, R.E.** 1991. Kalinin: an epithelium-specific basement membrane adhesion molecule that is a component of anchoring filaments. *J Cell Biol*. **114**:567-76.
  28. **Ryan, M.C., Tizard, R., VanDevanter, D.R., and Carter, W.G.** 1994. Cloning of the LamA3 gene encoding the alpha 3 chain of the adhesive ligand epiligrin. Expression in wound repair. *J Biol Chem*. **269**:22779-87.
  29. **Samantaray, S., Sharma, R., Chattopadhyaya, T.K., Gupta, S.D., Ralhan, R.** 2004. Increased expression of MMP-2 and MMP-9 in esophageal squamous cell carcinoma. *J Cancer Res Clin Oncol*. **130**:37-44.
  30. **Skyldberg, B., Salo, S., Eriksson, E., Aspenblad, U., Moberger, B., Tryggvason, K., and Auer, G.** 1999. Laminin-5 as a marker of invasiveness in cervical lesions. *J Natl Cancer Inst*. **91**:1882-7.
  31. **Soini, Y., Maatta, M., Salo, S., Tryggvason, K., Autio-Harmanen, H.** 1996. Expression of the laminin gamma 2 chain in pancreatic adenocarcinoma. *J Pathol*. **180**:290-4.
  32. **Sordat, I., Bosman, F.T., Dorta, G., Rousselle, P., Aberdam, D., Blum, A.L., and Sordat, B.** 1998. Differential expression of laminin-5 subunits and integrin receptors in human colorectal neoplasia. *J Pathol*. **185**:44-52.
  33. **Sordat, I., Rousselle, P., Chaubert, P., Petermann, O., Aberdam, D., Bosman, T.F., Sordat, B.** 2000. Tumor cell budding and laminin-5 expression in colorectal carcinoma can be modulated by the tissue micro-environment. *Int J Cancer*. **88**:708-17.
  34. **Stetler-Stevenson, W.G., Aznavoorian, S., and Liotta, L.A.** 1993. Tumor cell interactions with the extracellular matrix during invasion and metastasis. Review. *Annu Rev Cell Biol*. **9**:541-73.
  35. **Tournier, J.M., Polette, M., Hinnrasky, J., Beck, J., Werb, Z., and Basbaum, C.** 1994. Expression of gelatinase A, a mediator of extracellular matrix remodeling, by tracheal gland serous cells in culture and in vivo. *J Biol Chem*. **269**:25454-64.
  36. **Verrando, P., Pisani, A., and Ortonne, J-P.** 1988. The new basement membrane antigen recognized by the monoclonal antibody GB3 is a large size glycoprotein: modulation of its expression by retinoic acid. *Biochim Biophys Acta*. **942**:45-56.
  37. **Woessner, J.F. Jr.** 1991. Matrix metalloproteinases and their inhibitors in connective tissue remodeling. Review. *FASEB J*. **5**:2145-54.
  38. **Zhang, K., and Kramer, R.H.** 1996. Laminin 5 deposition promotes keratinocyte motility. *Exp Cell Res*. **227**:309-22.