

## Isolation of a Fission Yeast Mutant Cell Affected in MAP Kinase Signaling and Sterol Biosynthesis

KIWAMU IMAGAWA<sup>1</sup>, YUE FANG<sup>1,3</sup>, REIKO SUGIURA<sup>2</sup>,  
XIN ZHOU<sup>1</sup>, YAN MA<sup>1</sup>, and TAKAYOSHI KUNO<sup>1</sup>

<sup>1</sup>Division of Molecular Pharmacology and Pharmacogenomics Department of Biochemistry and Molecular Biology, Kobe University Graduate School of Medicine, Kobe, Japan;

<sup>2</sup>Laboratory of Molecular Pharmacogenomics, School of Pharmaceutical Sciences, Kinki University, Higashi-Osaka, Japan;

<sup>3</sup>Department of Pharmacology, China Medical University, Shenyang, China;

Received 27 January 2009/ Accepted 29 January 2009

**Key Words:** MAPK signaling, calcineurin, sterol, HMG-CoA reductase, farnesyltransferase

We have previously demonstrated that calcineurin and the Pmk1 MAP kinase pathway play an antagonistic role in Cl<sup>-</sup> homeostasis. Using this relationship, we screened for mutations that show *vic* (viable in the presence of immunosuppressant and chloride ion) phenotype and isolated a *vic6* mutant cell.

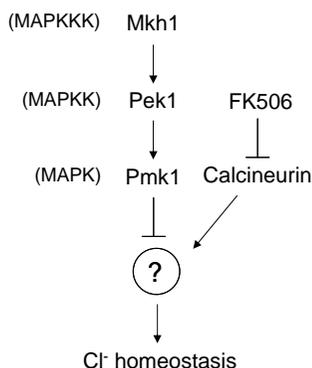
The *vic6* mutant cells also showed sensitivity to high temperature. Using this phenotype, we isolated *hmg1<sup>+</sup>* gene, encoding a HMG-CoA reductase. Consistently, the *vic6* mutant cells exhibited hypersensitivity to miconazole, an inhibitor of ergosterol biosynthesis and showed aberrant intracellular localization of filipin, suggesting that the mutant cells are affected in the sterol biosynthesis. In addition, overexpression of the *hmg1<sup>+</sup>* gene complemented the phenotype of *vic1-1/cpp1-v1* mutant cells, an allele of the gene encoding a farnesyltransferase, whereas overexpression of the *cpp1<sup>+</sup>* gene exacerbated the temperature-sensitive phenotype of the *vic6* mutant cells.

The mitogen-activated protein kinase (MAPK) signaling is one of the most important intracellular signaling that plays a crucial role in cell proliferation, cell differentiation, and cell cycle regulation (4,6,9,12). The Pmk1 MAPK, a homologue of the mammalian extracellular signal-regulated kinase (ERK)/MAPK, regulates cell morphology and cell integrity in fission yeast *Schizosaccharomyces pombe* (*S. pombe*) (15,18). Calcineurin, a Ca<sup>2+</sup>- and calmodulin-dependent protein phosphatase, is conserved from yeast to human (2,8,17) and is a molecular target for immunosuppressive drugs, such as cyclosporin A (CsA) and FK506 (7). These drugs induce their biological effects by forming an initial complex with cytosolic proteins termed immunophilins (cyclophilin and FKBP12). These drug-immunophilin complexes then bind to and inhibit calcineurin.

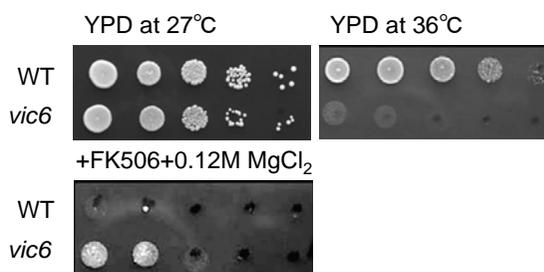
We have been studying basic cellular functions using fission yeast because this system is amenable to genetics and has many advantages in terms of relevance to higher systems. We have previously demonstrated that calcineurin plays an essential role in maintaining chloride ion homeostasis and acts antagonistically with the Pmk1 MAPK pathway (13,14 Figure.1). Based on this genetic interaction between calcineurin and Pmk1 MAPK, we screened for mutations that show *vic* (viable in the presence of immunosuppressant and chloride ion) phenotype and isolated the *vic* mutants (10). In the present study, we isolated and characterized *vic6* mutant cells and cloned *hmg1<sup>+</sup>* gene as a multicopy suppressor of the

## ISOLATION OF A FISSION YEAST MUTANT CELL

temperature-sensitive phenotype. Further analysis revealed that the mutated gene might be implicated in the sterol biosynthesis and that overexpression of *hmg1<sup>+</sup>* gene suppressed the phenotype of *cpl1-v1*, an allele of the *cpl1<sup>+</sup>* gene, encoding a farnesyltransferase. On the other hand, overexpression of the *cpl1<sup>+</sup>* gene exacerbated the temperature sensitivity of the *vic6* mutant cells suggesting a possible link between sterol biosynthesis and the MAPK pathway.



**Figure 1** Calcineurin and the Pmk1 MAPK pathway play an antagonistic role in Cl<sup>-</sup> homeostasis.



**Figure 2** The phenotypes of *vic6* cells. Wild-type and *vic6* mutant cells were dropped onto the plates as indicated and then incubated for 4 days at 27°C or 3 days at 36°C, respectively.

## MATERIALS AND METHODS

### Genetic Methods and Bioinformatics

Standard fission yeast molecular genetic methods were used except where noted (11, 15). Database searches were performed using the National Center for Biotechnology Information BLAST network service ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)) and the Sanger Center *S. pombe* data base search service ([www.sanger.ac.uk](http://www.sanger.ac.uk)).

### Microscopy and Miscellaneous Methods

Methods in light microscopy, such as fluorescence microscopy and differential interference contrast (DIC) microscopy, were performed as described (5). For staining of sterol, we used the fluorescent probe filipin, a polyene antibiotic that forms specific complexes with free 3- $\beta$ -hydroxysterols (3). Cells were grown to exponential phase in YPD medium at 27°C, and the filipin was added to the medium and cells were observed immediately.

## RESULTS AND DISCUSSION

### Isolation of *vic* Mutant

The *vic* mutants were isolated in a screen of cells that had been mutagenized with nitrosoguanidine as described previously (19). Mutants were spread on YPD plates to give ~1000 cells/plate, and the plates were incubated at 27°C for 4 days. The plates were then replica-plated at 27°C onto plates containing 0.5  $\mu$ g/ml FK506 and 0.2 M MgCl<sub>2</sub>. Mutants that grew in the plates were selected and designated as *vic* mutants. As shown in Table I, genes for several *vic* mutants have been identified. The *vic1<sup>+</sup>* gene encodes a  $\beta$  subunit of farnesyltransferase which acts upstream of Pmk1 MAPK signaling (10). Notably, many *vic*

mutants were resulted from mutation in the genes encoding members of Pmk1 MAPK signaling (Table I). These results are consistent with our hypothesis that calcineurin and the Pmk1 MAPK pathway play an antagonistic role in Cl<sup>-</sup> homeostasis.

**Table I.** Genes and gene products for *vic* mutants

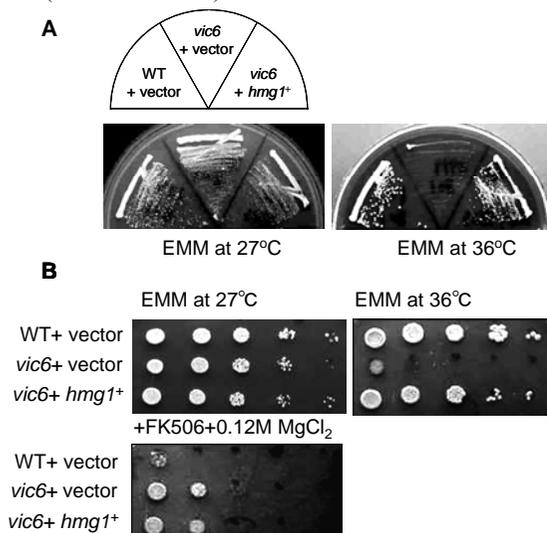
Mutant	Gene	Gene product
<i>vic1</i>	<i>cpp1</i> <sup>+</sup>	Farnesyltransferase β subunit
<i>vic8, vic13, vic16, vic18, vic19</i>	<i>pck2</i> <sup>+</sup>	Protein Kinase C
<i>vic9, vic14, vic17, vic21</i>	<i>pmk1</i> <sup>+</sup>	MAPK ( <i>S. pombe</i> MAPK)
<i>vic12, vic22</i>	<i>pek1</i> <sup>+</sup>	MAPKK ( <i>S. pombe</i> MEK)
<i>vic7, vic10, vic11, vic15, vic20, vic23</i>	<i>mkh1</i> <sup>+</sup>	MAPKKK ( <i>S. pombe</i> MEKK)

### Isolation and Characterization of the *vic6* Mutant Cell

The *vic6* mutant cell was also isolated from the screening for *vic* mutants as described above. As shown in Figure.2, the *vic6* mutant cells grew in the presence of FK506 and 0.12 M MgCl<sub>2</sub> at 27°C wherein wild-type cells failed to grow. The *vic6* mutant cells failed to grow at 36°C whereas wild-type cells grew normally.

### Isolation of the *hmg1*<sup>+</sup> Gene, Encoding a HMG-CoA Reductase, that Suppressed the Temperature-sensitive Phenotype of the *vic6* Mutant Cells

To clone the responsible gene by complementation, the temperature sensitivity of the *vic6* mutant cells was used. The *vic6* mutant cells were grown at 27°C and transformed with an *S. pombe* genomic DNA library constructed in the multicopy vector pDB248 (1). Leu<sup>+</sup> transformants were replica-plated onto YPD plates at 36°C, and the plasmid DNA was recovered from transformants that showed plasmid-dependent rescue. These plasmids complemented the temperature sensitivity. By DNA sequencing, one of the suppressing plasmid was identified to contain the *hmg1*<sup>+</sup> gene, which encodes a 1053-amino acids homolog of mammalian HMG-CoA reductase. The multicopy plasmid containing the *hmg1*<sup>+</sup> gene suppressed the temperature sensitive phenotype as shown in Figure.3A. However, overexpression of the *hmg1*<sup>+</sup> gene failed to suppress the *vic* phenotype as shown in Figure.3B. Consistently, further analysis suggested that the *vic6* mutant cell has at least two mutations (data not shown).



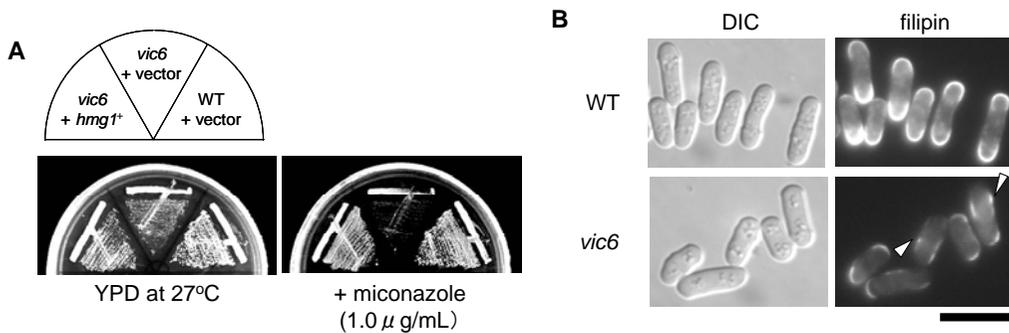
**Figure3** Overexpression of the *hmg1*<sup>+</sup> gene suppressed the temperature-sensitive phenotype of the *vic6* mutant cells, but not its *vic* phenotype. (A) Expression of *hmg1*<sup>+</sup> gene suppressed the temperature- sensitive phenotype of the *vic6* mutant cells. Cells were streaked onto EMM plates and then incubated for 4 days at 27°C or 3 days at 36°C, respectively. (B) The *hmg1*<sup>+</sup> gene failed to suppress the *vic* phenotype. Wild-type and *vic6* mutant cells were dropped onto the plates as indicated and then incubated for 4 days at 27°C or 3 days at 36°C, respectively.

## ISOLATION OF A FISSION YEAST MUTANT CELL

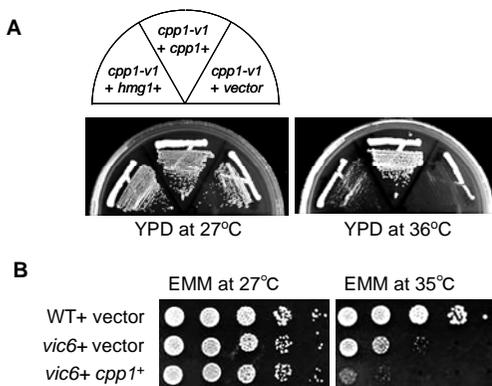
### The *vic6* Mutant Cells Were Hypersensitivity to Miconazole and Showed Aberrant Intracellular Localization of Filipin

The above results suggest that the sterol biosynthesis pathway may be defective in the *vic6* mutant cells. We then examined the effect of miconazole, an inhibitor of ergosterol biosynthesis. As shown in Figure.4A, the growth of the *vic6* mutant cells was significantly inhibited by miconazole as compared with that of wild-type cells. Next, we examined the cell morphology and the localization of the sterol, using the fluorescent probe filipin, a polyene antibiotic that forms specific complexes with free 3- $\beta$ -hydroxysterols. The sterol is the final product for sterol biosynthesis pathway. As shown in Figure.4B, in wild-type cells filipin is enriched in the plasma membrane at the growing cell tips and as reported by Wachtler *et al.* (16). On the other hand, filipin fluorescence was weakly observed at the plasma membrane outside the growing cell tips of the *vic6* mutant cells (arrowheads).

Taken together with the result that the *hmg1<sup>+</sup>* gene is isolated as a multicopy suppressor, these results suggest that sterol biosynthesis and/or sterol transport was defective in the *vic6* mutant cells.



**Figure4** The *vic6* mutant cells showed hypersensitivity to miconazole and aberrant localization of filipin. (A) The *vic6* mutant cells were hypersensitive to miconazole, an inhibitor of ergosterol biosynthesis. (B) Cell morphology and intracellular localization of filipin. DIC; differential interference contrast. Arrowheads indicate filipin fluorescence that was weakly observed at the plasma membrane outside the growing cell tips. Bar, 10  $\mu$ m.



**Figure5** Genetic interaction between the *hmg1<sup>+</sup>* gene and the *cpp1<sup>+</sup>* gene. (A) Overexpression of the *hmg1<sup>+</sup>* gene suppressed the temperature sensitive phenotype of the *vic1-1/cpp1-v1* mutant. Cells were streaked onto YPD plates and then incubated for 4 days at 27°C or 3 days at 36°C, respectively. (B) Overexpression of the *cpp1<sup>+</sup>* gene exacerbated the temperature sensitivity of the *vic6* mutant cells. Wild-type and the *vic6* mutant cells were dropped onto the plates and then incubated for 4 days at 27°C or 3 days at 35°C, respectively.

### The *hmg1*<sup>+</sup> Gene Complemented the Temperature Sensitivity of *vic1-1/cpp1-v1* Mutant

Next, we examined the genetic interaction between the *hmg1*<sup>+</sup> gene and the responsible gene for *vic1* mutant, *cpp1*<sup>+</sup>, encoding a  $\beta$  subunit of farnesyltransferase (Table I). Overexpression of *hmg1*<sup>+</sup> gene complemented the temperature-sensitive phenotype of *vic1-1/cpp1-v1* mutant (Figure.5A), but not that of *cpp1* deletion (data not shown). The farnesyltransferase is located downstream from HMG-CoA reductase in the sterol biosynthesis pathway and catalyzes the posttranslational modification of Ras family proteins. Cpp1 functions upstream Pmk1 MAPK signaling through Rho2 regulation (10). These results suggest that overexpression of the *hmg1*<sup>+</sup> gene increased the intermediates of the sterol biosynthesis, including farnesyl diphosphate, which is used as the source for farnesylation. This result prompted us to investigate whether overexpression of the *cpp1*<sup>+</sup> gene affects the phenotype of *vic6* mutant cells. Interestingly, overexpression of the *cpp1*<sup>+</sup> gene resulted in exacerbation of temperature-sensitive phenotype of the *vic6* mutant cells (Figure.5B). These results suggest a possible relationship between MAPK signaling and sterol biosynthesis pathway in fission yeast. Presumably, overexpression of the *cpp1*<sup>+</sup> gene in the *vic6* mutant cells may promote overconsumption of farnesyl diphosphate and shortage of the downstream products in the pathway, thereby exacerbating the temperature sensitivity of the mutant. Consistently, *vic6* mutant cells showed hypersensitivity to miconazole and aberrant localization of filipin, suggesting that sterol biosynthesis is decreased in the *vic6* mutant cells.

### ACKNOWLEDGEMENTS

We thank Susie O. Sio for critical reading of the manuscript. This work was supported by 21st Century COE Program, Global COE Program and research grants from the Ministry of Education, Culture, Sports, Science and Technology of Japan.

### REFERENCES

1. **Beach, D., Piper, M., and Nurse, P.** 1982. Construction of a *Schizosaccharomyces pombe* gene bank in a yeast bacterial shuttle vector and its use to isolate genes by complementation. *Mol. Gen. Genet* **187**: 326-329.
2. **Cyert, M.S., Kunisawa, R., Kaim, D., and Thorner, J.** 1991 Yeast has homologs (*CNA1* and *CNA2* gene products) of mammalian calcineurin, a calmodulin-regulated phosphoprotein phosphatase. *Proc Natl Acad Sci USA* **88**:7376-7380.
3. **Drabikowski, W., Lagwinska, E., and Sarzala, M.G.** 1973. Filipin as a fluorescent probe for the location of cholesterol in the membranes of fragmented sarcoplasmic reticulum. *Biochim. Biophys. Acta* **291**:61-70.
4. **Herskowitz, I.** 1995. MAP kinase pathways in yeast: for mating and more. *Cell* **80**: 187-197.
5. **Kita, A., Sugiura, R., Shoji, H., He, Y., Deng, L., Lu, Y., Sio, S. O., Takegawa, K., Sakaue, M., Shuntoh, H., and Kuno, T.** 2004. Loss of Apm1, the micro1 subunit of the clathrin-associated adaptor-protein-1 complex, causes distinct phenotypes and synthetic lethality with calcineurin deletion in fission yeast. *Mol. Biol. Cell* **15**:2920-2931.
6. **Levin, D.E., and Errede, B.** 1995. The proliferation of MAP kinase signaling pathways in yeast. *Curr. Opin. Cell Biol* **7**:197-202.

## ISOLATION OF A FISSION YEAST MUTANT CELL

7. **Liu, J., Farmer, J.D. Jr., Lane, W.S., Friedman, J., Weissman, I., and Schreiber, S.L.** 1991a. Calcineurin is a common target of cyclophilin-cyclosporin A and FKBP-FK506 complexes. *Cell* **66**:807-815.
8. **Liu, Y., Ishii, S., Tokai, M., Tsutsumi, H., Ohki, O., Akada, R., Tanaka, K., Tsuchiya, E., Fukui, S., and Miyakawa, T.** 1991b. The *Saccharomyces cerevisiae* genes (*CMP1* and *CMP2*) encoding calmodulin-binding proteins homologous to the catalytic subunit of mammalian protein phosphatase 2B. *Mol.Gen.Genet.* **227**, 52-59.
9. **Marshall, C. J.** 1994. MAP kinase kinase kinase, MAP kinase kinase and MAP kinase. *Curr. Opin. Genet. Dev* **4**:82-89.
10. **Ma, Y., Kuno, T., Kita, A., Asayama, Y., and Sugiura, R.** 2006. Rho2 is a target of the Farnesyltransferase Cpp1 and acts upstream of Pmk1 mitogen-activated protein kinase signaling in Fission Yeast. *Mol. Biol. Cell*, Dec; **17**: 5028-5037.
11. **Moreno, S., Klar, A., and Nurse, P.** 1991. Molecular genetic analysis of fission yeast *Schizosaccharomyces pombe*. *Methods Enzymol.* **194**:795-823.
12. **Nishida, E., and Gotoh, Y.** 1993. The MAP kinase cascade is essential for diverse signal transduction pathways. *Trends Biochem. Sci* **18**:128-131.
13. **Sugiura, R., Toda, T., Dhut, S., Shuntoh, H., and Kuno, T.** 1999 The MAPK kinase Pck1 acts as a phosphorylation-dependent molecular switch. *Nature.* **399**:479-483.
14. **Sugiura, R., Toda, T., Shuntoh, H., Yanagida, M., and Kuno, T.** 1998 *pmp1<sup>+</sup>*, a suppressor of calcineurin deficiency, encodes a novel MAP kinase phosphatase in fission yeast. *EMBO J.* **17**:140-148.
15. **Toda, T., Dhut, S., Superti-Furga, G., Gotoh, Y., Nishida, E., Sugiura, R., and Kuno, T.** 1996. The fission yeast *pmp1<sup>+</sup>* gene encodes a novel mitogen-activated protein kinase homolog which regulates cell integrity and functions coordinately with the protein kinase C pathway. *Mol. Cell Biol.* **16**:6752-6764.
16. **Wachtler, V., Rajagopalan, S., and Balasubramanian, M.K.** 2003. Sterol-rich plasma membrane domains in the fission yeast *Schizosaccharomyces pombe*. *J. Cell Sci.* **116**:867-874.
17. **Yoshida, T., Toda, T., and Yanagida, M.** 1994. A calcineurin-like gene *ppb1<sup>+</sup>* in fission yeast: mutant defects in cytokinesis, cell polarity, mating and spindle pole body positioning. *J.Cell Sci.* **107 (Pt 7)** :1725-1735
18. **Zaitsevskaya-Carter, T., and Cooper, J.A.** 1997. Spm1, a stress-activated MAP kinase that regulates morphogenesis in *S. pombe*. *EMBO J* **16**:1318-1331.
19. **Zhang, Y., Sugiura, R., Lu, Y., Asami, M., Maeda, T., Itoh, T., Takenawa, T., Shuntoh, H., and Kuno, T.** 2000 Phosphatidylinositol-4-phosphate 5-kinase Its3 and calcineurin Ppb1 coordinately regulate cytokinesis in fission yeast. *J. Biol. Chem. Nov;* **275 (45)**:35600-6.